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## **Gene Section**

**Short Communication** 

# ERVE-1 (endogenous retrovirus group E member 1)

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#### Abstract

Endogenous retrovirus group E member 1, alias ERVE-1, is an integrated retroviral element of the human genome. Still poorly characterized, it maps on Chromosome 17 and it is reported for multiple sclerosis and systemic lupus erythematosus.

#### Keywords

Endogenous retrovirus; ERVs; ERVE-1

## **Identity**

#### Other names:

ERVE1; endogenous retroviral sequence E, 1; endogenous retrovirus group E, member 1; HERV-E1; HERVE1; TCONS\_00025599; Lnc-IFT20-4; HERVE a

**HGNC (Hugo):** ERVE-1 **Location:** 17q11.2

#### Local order

Starts at 28,232,590 and ends at 28,235,281 bp from gter, reverse strand (GRCh38/hg38)

### **DNA/RNA**

Endogenous retrovirus group E member 1 (ERVE-1) is an integrated retroviral element of the human genome. It is known that human endogenous retroviruses (ERVs) and ERV-like sequences are part of the repetitive portions of the human genome and are remnants of infections of former exogenous retroviruses. They comprise about 8% of the human genome (Landers et al, 2001) and include solitary

long terminal repeats (LTRs), non-retroviral sequences flanked by LTRs, and LTRs-like sequences (Mayer et al. 2011).

ERVs are divided into various groups. ERVE-1 is member 1 of the group E of endogenous retroviruses and it maps on Chromosome 17, specifically on 17q11.2 (Campos-Sánchez and Sandoval-Carvajal, 2018; Taruscio et al, 2002). In addition, it starts at 28,232,590 and ends at 28,235,281 bp from qter (reverse strand). It is present in the Ensembl database with a sequence named ENSG00000267259. ERVE-1 DNA sequence shows a strong promoter sequence site located at +0.8 kb from the gene transcription start site

#### **Transcription**

Usually, human ERVs are silenced epigenetically and are not transcribed (Kewitz and Staege, 2013), however, they are found to be transcribed in particular situations, such as under pathological conditions, and usually their transcription is initiated by promoter sequences within the proviral LTRs (Mayer et al, 2011).

ERVE-1 mRNA is 2,691 bp long with a reference sequence reported in GeneBank as BC037342. It is also present in the Ensembl database with a sequence named ERVE-1-201 (ENST00000592016.1) and it is considered to be a long non-coding RNA (lncRNA). Abundant ERVE-1 transcript was found in normal pancreas and thyroid tissues (Shiroma et al, 2001) and this suggests that it may play a role in their physiologic functions.

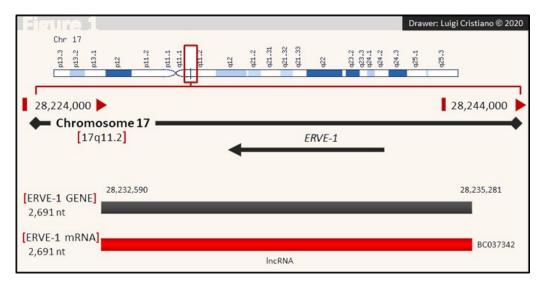


Figure. 1. ERVE-1 gene. The figure shows the locus on chromosome 17 of the ERVE-1 gene, its DNA and mRNA (reworked from https://www.ncbi.nlm.nih.gov/gene; http://grch37.ensembl.org; www.genecards.org)

In addition, the long terminal repeat (LTR)-derived promoter of ERVE-1 was found hypomethylated in placental cells whereas it was detected hypermethylated in blood cells (Ruebner et al, 2013; Reiss et al, 2007). This suggests that ERVE-1 could be transcribed in this tissue compared to others in which it is epigenetically silenced.

#### **Pseudogene**

No observed pseudogenes for the ERVE-1 sequence.

#### **Protein**

#### Note

It is well-known that human ERVs and ERV-like sequences no longer encode proteins because of the accumulation of nonsense mutations as a consequence of their ancient incorporation into the genome of the host (Mayer et al, 2011). For ERVE-1 sequence there is no protein reported.

## Implicated in

Expression of ERVs is usually switched off (Kewitz and Staege, 2013) but the expression of ERVs and ERV-like sequences have been reported for a large number of human diseases, including cancer, however, their exact involvement and mechanism of action remain to be clarified (Mayer et al, 2011). ERVE-1 is poorly studied, however, it was reported for multiple sclerosis (MS) and Systemic lupus erythematosus (SLE).

#### Multiple sclerosis (MS)

ERVE-1 could have biological functions in the pathogenesis of multiple sclerosis (MS) (Kim et al, 2008; Antony et al, 2007), however, as the disease is very complex, further studies are needed to understand its contribution.

#### Systemic lupus erythematosus (SLE)

The expression level of ERVE-1 mRNA is found higher in lupus CD4+ T cells respect to healthy controls while the methylation of the LTR-derived promoter of ERVE-1 is found low in these patients (Suntsova et al, 2015; Wu et al, 2015). These data suggest that ERVE-1 is positively correlated with systemic lupus erythematosus (SLE) disease and that it is involved in its development (Wu et al, 2015).

#### References

Antony JM, Zhu Y, Izad M, Warren KG, Vodjgani M, Mallet F, Power C. Comparative expression of human endogenous retrovirus-W genes in multiple sclerosis AIDS Res Hum Retroviruses 2007 Oct;23(10):1251-6.

Kewitz S, Staege MS. Expression and Regulation of the Endogenous Retrovirus 3 in Hodgkin's Lymphoma Cells Front Oncol 2013 Jul 10:3:179.

Kim HS, Ahn K, Kim DS. Quantitative expression of the HERV-W env gene in human tissues Arch Virol 2008;153(8):1587-91.

Lander ES, Linton LM, Birren B, Nusbaum C, Zody MC, Baldwin J, Devon K, Dewar K, Doyle M, FitzHugh W, Funke R, Gage D, Harris K, Heaford A, Howland J, Kann L, Lehoczky J, LeVine R, McEwan P, McKernan K, Meldrim J, Mesirov JP, Miranda C, Morris W, Naylor J, Raymond C, Rosetti M, Santos R, Sheridan A, Sougnez C, Stange-Thomann Y, Stojanovic N, Subramanian A. Wyman D, Rogers J, Sulston J, Ainscough R, Beck S, Bentley D, Burton J, Clee C, Carter N, Coulson A, Deadman R, Deloukas P, Dunham A, Dunham I, Durbin R, French L, Grafham D, Gregory S, Hubbard T, Humphray S, Hunt A, Jones M, Lloyd C, McMurray A, Matthews L, Mercer S, Milne S, Mullikin JC, Mungall A, Plumb R, Ross M, Shownkeen R, Sims S, Waterston RH, Wilson RK, Hillier LW, McPherson JD, Marra MA, Mardis ER, Fulton LA, Chinwalla AT, Pepin KH, Gish WR, Chissoe SL, Wendl MC, Delehaunty KD, Miner TL, Delehaunty A, Kramer JB, Cook LL, Fulton RS, Johnson DL, Minx PJ, Clifton SW. Hawkins T, Branscomb E, Predki P, Richardson P, Wenning S, Slezak T, Doggett N, Cheng JF, Olsen A,

Lucas S, Elkin C, Uberbacher E, Frazier M, Gibbs RA, Muzny DM, Scherer SE, Bouck JB, Sodergren EJ, Worley KC. Rives CM, Gorrell JH, Metzker ML, Naylor SL, Kucherlapati RS, Nelson DL, Weinstock GM, Sakaki Y, Fujiyama A, Hattori M, Yada T, Toyoda A, Itoh T, Kawagoe C, Watanabe H, Totoki Y, Taylor T, Weissenbach J, Heilig R, Saurin W, Artiguenave F, Brottier P, Bruls T, Pelletier E, Robert C, Wincker P, Smith DR, Doucette-Stamm L, Rubenfield M, Weinstock K, Lee HM, Dubois J, Rosenthal A, Platzer M, Nyakatura G, Taudien S, Rump A, Yang H, Yu J, Wang J, Huang G, Gu J, Hood L, Rowen L, Madan A, Qin S, Davis RW, Federspiel NA, Abola AP, Proctor MJ, Myers RM, Schmutz J, Dickson M, Grimwood J, Cox DR, Olson MV, Kaul R, Raymond C, Shimizu N, Kawasaki K, Minoshima S, Evans GA, Athanasiou M, Schultz R, Roe BA, Chen F, Pan H, Ramser J, Lehrach H, Reinhardt R, McCombie WR, de la Bastide M, Dedhia N, Blöcker H, Hornischer K, Nordsiek G, Agarwala R, Aravind L, Bailey JA, Bateman A, Batzoglou S, Birney E, Bork P, Brown DG, Burge CB, Cerutti L, Chen HC, Church D, Clamp M, Copley RR, Doerks T, Eddy SR, Eichler EE, Furey TS, Galagan J, Gilbert JG, Harmon C, Hayashizaki Y, Haussler D, Hermjakob H, Hokamp K, Jang W, Johnson LS, Jones TA, Kasif S, Kaspryzk A, Kennedy S, Kent WJ, Kitts P, Koonin EV, Korf I, Kulp D, Lancet D, Lowe TM, McLysaght A, Mikkelsen T, Moran JV, Mulder N, Pollara VJ, Ponting CP, Schuler G, Schultz J, Slater G, Smit AF, Stupka E, Szustakowki J, Thierry-Mieg D, Thierry-Mieg J, Wagner L, Wallis J, Wheeler R, Williams A, Wolf YI, Wolfe KH, Yang SP, Yeh RF, Collins F, Guyer MS, Peterson J, Felsenfeld A, Wetterstrand KA, Patrinos A, Morgan MJ, de Jong P, Catanese JJ, Osoegawa K, Shizuya H, Choi S, Chen YJ, Szustakowki J,. Initial sequencing and analysis of the human genome Nature 2001 Feb 15;409(6822):860-921.

Mayer J, Blomberg J, Seal RL. A revised nomenclature for transcribed human endogenous retroviral loci Mob DNA

2011 May 4;2(1):7.

Reiss D, Zhang Y, Mager DL. Widely variable endogenous retroviral methylation levels in human placenta Nucleic Acids Res 2007;35(14):4743-54.

Ruebner M, Strissel PL, Ekici AB, Stiegler E, Dammer U, Goecke TW, Faschingbauer F, Fahlbusch FB, Beckmann MW, Strick R. Reduced syncytin-1 expression levels in placental syndromes correlates with epigenetic hypermethylation of the ERVW-1 promoter region PLoS One 2013;8(2):e56145.

Shiroma T, Sugimoto J, Oda T, Jinno Y, Kanaya F. Search for active endogenous retroviruses: identification and characterization of a HERV-E gene that is expressed in the pancreas and thyroid J Hum Genet 2001;46(11):619-25.

Suntsova M, Garazha A, Ivanova A, Kaminsky D, Zhavoronkov A, Buzdin A. Molecular functions of human endogenous retroviruses in health and disease Cell Mol Life Sci 2015 Oct;72(19):3653-75.

Taruscio D, Floridia G, Zoraqi GK, Mantovani A, Falbo V. Organization and integration sites in the human genome of endogenous retroviral sequences belonging to HERV-E family Mamm Genome 2002 Apr;13(4):216-22.

Wu Z, Mei X, Zhao D, Sun Y, Song J, Pan W, Shi W. DNA methylation modulates HERV-E expression in CD4+ T cells from systemic lupus erythematosus patients J Dermatol Sci 2015 Feb;77(2):110-6.

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